

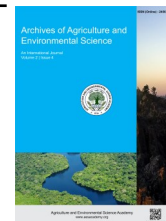


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REVIEW ARTICLE



## A review on the impact of commonly used pesticides on the biology of earthworms

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### ABSTRACT

Earthworms are considered important bio-indicators of chemical contamination in the soil ecosystem. Being an important biotic factor of soil ecosystem, earthworms play a vital role in the functioning of soil ecosystems and maintenance of soil fertility. The present review encompasses the diverse effects of chemical contaminants like pesticides on earthworm biology, considering both direct toxicity and indirect impacts on ecosystem functions. Through a comprehensive review of existing literature, we assess the varying impacts of different classes of pesticides on earthworms. Several studies included in this review shed light on how pesticide exposure affects earthworm behaviour, reproduction, regenerative capacity, histology, gut microbial diversity, and nutrient transition, among other adverse effects, which consequently affect the soil ecosystem dynamics. Furthermore, we discuss the implications of these findings for agricultural practices, soil health, and biodiversity conservation. This study discusses the impact of pesticides on different facets of earthworm biology and emphasizes the necessity of sustainable pest management strategies to maintain the productivity and adaptability of ecosystems by enhancing our understanding of the complex interactions that occur between soil organisms, like earthworms, and foreign chemicals, or xenobiotics, like pesticides.

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### INTRODUCTION

Earthworms are distributed worldwide, and their population is considered to make up approximately 8% of the total biomass of the soil organisms (Sinha *et al.*, 2013). It is a tube-shaped, long, cylindrical, segmented soil-dwelling creature belonging to the class Oligochaeta of Phylum Annelida. These soil organisms play various significant roles in maintenance and development of the nutritional value and quality of soil by turning organic waste and biodegradable material into nutrient-rich worm manure (Bardiya & Dixit, 2024). The cast produced by earthworms have higher organic content than that of the surrounding soil in which earthworms remain present (Bhadauria & Saxena, 2010). Being a vital biotic component of the soil ecosystem, earthworms significantly contribute to enhancing soil fertility and maintaining the soil ecosystem dynamics by increasing nutrient availability

through their potential role in organic matter mineralization and decomposition (Aina, 1984; Edwards & Bohlen, 1996; Abdul & Bouché, 1997; Brown *et al.*, 2003). Therefore, any factor that can adversely affect the growth, survival, biological parameters, or ecological functioning of earthworms might have a detrimental impact on soil quality and soil ecosystem dynamics. The rapid increase in human population has a mandatory demand for a rapid and corresponding increase in crop production, which has resulted in the irrational use of different chemical agents like pesticides for crop protection and better crop yield. With the onset of Green revolution in 1960s the use of synthetic chemical agents like pesticides has increased rapidly across the globe including India (Alavanja, 2009). The total amount of pesticides used worldwide is approximately 3.39 million tonnes, whereas in India it is approximately 61,702 tonnes (Reddy & Mathur, 2024). In addition, Reddy & Mathur (2024) noted that

while pesticide use in India has increased by about 46% overall, there has been a notable spike in use in a few states, including Jharkhand (an increase of 833%), Chhattisgarh (247%), Andhra Pradesh (265%), and Maharashtra (253%). However, these chemical agents can also have adverse impact on beneficial and non-target soil organisms, most notably earthworms (Gowri & Thangaraj, 2020). The earthworms are considered significant bio-indicators of contamination of soil by undesirable agents, including pesticides (Edwards & Thompson, 1973; Greig-Smith et al., 1992). There are a number of pesticides that are extensively used in agriculture, such as imidacloprid, benomyl, and metribuzin among many others which are reported to have a harmful or adverse impact on earthworms (Astaykina et al., 2022).

Pesticides are categorized into four primary groups based on their chemical composition: Carbamates, Pyrethrin and Pyrethroids, Organochlorines, and Organophosphorus (Buchel, 1983). Pesticides reduce biodiversity even at concentrations that don't go above the Maximum Residue Levels (MRLs) (Hole et al., 2005). In addition to their role in improving crop yield, pesticides can adversely affect the environment in several ways. According to Reinecke & Reinecke (2007), the usage of organophosphate insecticides has caused significant changes in earthworm population densities in South Africa. Another study from Mahmood et al. (2016) has reported that excessive pesticide use may result in the eradication of biodiversity. The target animal (or organism) and chemical structure (organic, inorganic, synthetic, and biological compounds) are used to categorize pesticides (Amenyogbe et al., 2021). Implementing appropriate corrective actions contributes to fewer cases of pesticide poisoning and other health problems associated with pesticide use (Rani et al., 2021). Persistent pesticides degrade slowly and can linger in the environment for extended periods and the soil conditions can also impact pesticide transport (Anjum et al., 2017). Previous studies have found that class I and II pesticides (WHO, 2005), extremely to moderately hazardous and banned in other countries, are still available in India (Bond et al., 2009). Many studies have been carried out in the last few decades on how different pesticides affect different aspects of earthworm biology, and only a small number of these pesticides are no longer in use. The employment of novel chemical combinations as pesticides, however, is possible everywhere in the world, especially in India. Understanding the effects of commonly used pesticides on earthworm biology and ecology is crucial, especially considering the significant roles that earthworms play in soil ecology and the widespread use of pesticides. It is quite evident that a wide range of pesticides are employed in agricultural fields, and the majority of these pesticides have not been tested for their effects on earthworm biology or ecology. As the survival of earthworms and its contributions in soil ecosystem have many facets therefore it is necessary to understand the impact of any pesticide in all such aspects which can affect the survival and ecological role of earthworms. Wang et al. (2012) have studied almost 45 different types of pesticide compositions on a

single earthworm species, *Eisenia fetida*, and it can be viewed in correlation with different types of earthworm species like epigeic, anecic, and endogeic, as well as a significant biodiversity of different earthworm species. Additional research in this area is necessary to better understand earthworm-pesticide interactions because of the wide range of earthworm species, the various pesticide compositions in use, and the need to understand how each pesticide affects various aspects of earthworm biology. Therefore, a comprehensive understanding of the impact of chemical agents on soil ecosystem dynamics and soil ecology is essential. The present review encompasses the scientific reports and evaluation of the impact of some commonly used pesticides on some selected parameters of biology and ecological role of earthworms and might open the ways of new or further research in this field so that the use of any kind of chemical agent to increase crop yields will be made in a sustainable way to protect the soil faunal diversity and ecology, especially the earthworms.

## METHODOLOGY

The primary objective of the present study is to evaluate the inferences drawn from existing scientific research works in the field of pesticide-earthworm interactions and to assess the gaps in understanding the impact of varieties of pesticides on diverse facets of earthworm biology, from molecular to ecological levels. We started our work by collecting the data on pesticide use from official sources like Food and Agriculture Organization Statistics (FAOSTAT) and a few Indian government publications. Later on, we went through the results of previous research works on pesticide-earthworm interactions through a survey of available and current scientific literature. We then categorized the major facets of earthworm biology that have been worked out to evaluate the impact of selected pesticides and then designed our work to focus on the selected aspects of earthworm biology, such as growth and development, reproduction, regeneration, oxidative stress, gut microbial diversity, and histological impacts, for evaluation of the impact of different pesticides. From the available scientific works, it was quite evident that during the past few decades till date, a number of chemical combinations have been utilized as pesticides for crop protection; on the contrary, there are different ecological groups and varieties of different species of earthworms. Neither each of the pesticides is being assessed for its impact on all the significant aspects of earthworm biology, nor are all the different varieties of earthworm species being evaluated to see the impact of a single pesticide. Further, the impact of a single pesticide on different ecological groups of earthworms, such as epigeic, anecic, and endogeic, found in the same geographical area, is also not worked out. The lack of a comprehensive and comparative analysis of the impact of widely used pesticide chemical combinations on different facets of earthworm biology is existing, and therefore the present study has been undertaken to address the available scientific knowledge and the gaps in the field of pesticide-earthworm interactions.

### Effects of some commonly used pesticides on earthworms

Pesticides have the potential to negatively affect earthworms at various organizational and biological levels. These effects may include behavioral modifications, altered growth and development, altered fertility and reproduction, increased mortality, changes in metabolic features such as enzyme function, and even changes in gene expression (Zhang et al., 2013; Velki et al., 2014; Velki & Ešimović, 2015; Liu et al., 2017, 2018). The potential impact of pesticides on earthworms depends upon various factors such as earthworm species, type and concentration of contaminants, soil properties, etc. (Rodriguez-Campos et al., 2014). The epigeic and anecic earthworm species are found in the upper soil layers near the surface, whereas the endogeic species remain present in the inner soil strata, and therefore the

epigeic and anecic species are more prone to facing the adverse effects of pesticides since the pesticides are used to spray on the soil surface (Singh et al., 2016). Several studies have been conducted to evaluate the impact of commonly used pesticides on a range of biological parameters of various earthworm species, including histopathological effects, enzyme activities and oxidative stress assessment, growth and reproduction (including gamete viability, cocoon production, etc.), gut microbial diversity, cast quality and production, cytotoxic activities, behavioural changes like burrowing activities, etc. (Datta et al., 2016). Some of the scientific reports of commonly used pesticides and their impact on different parameters of earthworm species are listed in Table 1.

**Table 1.** Commonly used pesticides and their impact on different biological parameters of selected earthworm species.

S. No.	Pesticide	Earthworm species	Effect	Reference
1.	Carbaryl and Dieldrin	<i>Eisenia fetida</i>	Growth and cocoon production inhibited.	Neuhauser & Callahan (1990)
2.	Benomyl	<i>Eisenia fetida</i>	Sperm were undeveloped or had abnormal acrosome formation.	Sorour & Larink (2001)
3.	Carbaryl	<i>Eisenia fetida</i>	Cholinesterase (ChE) inhibition was observed even at low doses.	Ribera et al. (2001)
4.	Butachlor, Malathion and Carbofuran	<i>Drawida willsi</i>	Butachlor did not affect AChE activity. Maximum AChE inhibition occurred after 9 days of malathion treatment and 12 days of Carbofuran exposure.	Booth & O'Halloran (2001)
5.	Chlorpyrifos, Atrazine, Cyanazine	<i>Eisenia fetida</i>	Chlorpyrifos binary combinations with Atrazine and Cyanazine were more harmful than additives.	Lydy & Linck (2003)
6.	Endosulphan and Aldicarb	<i>Lumbricus terrestris</i>	Weight loss and reduced growth rate. Compared to Endosulfan, Aldicarb is more hazardous and toxic.	Mosleh et al. (2003)
7.	Azodrin	<i>Eisenia fetida</i>	Dose-dependent suppression of AChE activity. AChE activity is associated with morphological damage.	Rao & Kavitha (2004)
8.	Butachlor, Malathion and Carbofuran	<i>Drawida willsi</i>	No variation in AChE activity when exposed to Butachlor; maximum AChE inhibition was observed (41% and 46%) after 9 days when exposed to malathion and after 12 days of Carbofuran exposure (54% and 62.9%).	Panda & Sahu (2004)
9.	Malathion	<i>Eisenia fetida</i>	Significant loss of body mass and decline in sperm viability.	Navarro & Obregon (2005)
10.	Imidacloprid	<i>Aporrectodea nocturna</i> and <i>Allolobophora icterica</i>	The LC50 for <i>Aporrectodea nocturna</i> and <i>Allolobophora icterica</i> was 2-4 mg kg <sup>-1</sup> dry soil. Additionally, considerable weight loss was noticed.	Capowiez et al. (2005)
11.	Acetochlor	<i>Eisenia fetida</i>	The growth and number of juveniles within each cocoon were greatly impacted.	Xiao et al. (2006)
12.	Carbofuran	<i>Eisenia fetida</i>	Total Cholinesterase Activity and protein content both increased at low levels, and vice versa. When the concentration of Carbofuran rose, SOD decreased and vice versa.	Ling (2006)
13.	Imidacloprid	<i>Aporrectodea nocturna</i> and <i>Allolobophora icterica</i>	Both species disrupted the continuity of their burrow systems. There was a decrease in gas diffusion through the soil cores of <i>Aporrectodea nocturna</i> , while there was no change in gas diffusion in the soil cores of <i>Allolobophora icterica</i> .	Capowiez & Berard (2006)
14.	Benomyl	<i>Eisenia fetida</i>	The toxicity of Benomyl was lower in tropical than temperate artificial soils.	Rombke et al. (2007)
15.	Carbendazim, Dimethoate, and Glyphosate	<i>Eisenia fetida</i>	Both independently and in combination are harmful to reproduction and development.	Yasmin & D'Souza (2007)
16.	Chlorpyrifos	<i>Eisenia fetida</i>	Adverse effects on growth and development	Zhou et al. (2007)
16.	Lindane and deltamethrin	<i>Eisenia fetida</i>	Lindane was found to be more harmful than deltamethrin. There were significant effects on growth and cellulase activity.	Shi et al. (2007)
17.	Glyphosate and Chlorpyrifos	<i>Eisenia fetida andrei</i>	Glyphosate decreased cocoon production. The NRRT (Neutral red retention test) and Comet assays demonstrated changes at the subcellular level.	Casabé et al. (2007)
18.	Carbaryl	<i>Eisenia fetida andrei</i>	AChE activity was seen to be inhibited dose-dependently.	Gambi et al. (2007)
19.	Chlorpyrifos and Azinphos methyl	<i>Aporrectodea caliginosa</i>	Inhibition of ChE.	Reinecke & Reinecke (2007)
20.	Cypermethrin	<i>Eisenia fetida</i>	Notable decrease in the production of cocoons. Juveniles were more sensitive than adults.	Shi-ping et al. (2008)

Table 1. Contd.....

21.	Cypermethrin	<i>Eisenia fetida Andrei</i>	Impacts the reproductive functions and reduction in cocoon production. Juveniles more sensitive than adults.	Ahmad et al. (2012); Lu et al. (2021)
21.	Butachlor	<i>Eisenia fetida</i>	Using the chloragogen tissue's reserve energy results in less biomass and cocoons.	Gobi & Gunasekaran (2010)
22.	Imidacloprid	<i>Lumbricus terrestris</i> and <i>Aporrectodea caliginosa</i>	Body mass loss and increased cast formation at higher concentrations.	Dittbrenner et al. (2010)
23.	Chlorpyrifos, Carbofuran and Mancozeb	<i>Perionyx excavatus</i>	The decline in toxicity: Carbofuran > Chlorpyrifos > Mancozeb	De Silva et al. (2010)
24.	Cypermethrin, Endosulfan, Carbaryl, Chlorpyrifos, Aldicarb, Monocrotophos	<i>Perionyx excavatus</i>	Toxicity order Monocrotophos > Aldicarb > Chlorpyrifos > Carbaryl > Cypermethrin > Endosulfan	Gupta et al. (2010)
25.	Imidacloprid	<i>Pheretima posthuma</i>	Protein concentration increased in the clitellum, some proteins were inhibited in the head, and there was no change seen in abdomen.	Faheem & Khan (2010)
26.	R-metalaxyl and rac-metalaxyl	<i>Eisenia fetida</i>	Metalaxyl bioaccumulation was found to be enantioselective in earthworms, with preferential accumulation of the Senantiomer.	Xu et al. (2011)
27.	Imidacloprid	<i>Lumbricus terrestris</i> and <i>Aporrectodea caliginosa</i>	<i>Aporrectodea caliginosa</i> exhibits burrowing effects even at lower concentrations, but <i>Lumbricus terrestris</i> exhibits burrowing effects only at greater concentrations.	Dittbrenner et al. (2011)
28.	Mixture of Ni and Chlorpyrifos	<i>Lumbricoid</i>	The combination of Ni and chlorpyrifos is harmful to earthworms. Worms store Ni and chlorpyrifos in their tissues.	Lister et al. (2011)
29.	Chlorpyrifos and Fenvalerate	<i>Eisenia fetida</i>	Cellulase and SOD activity were inhibited, while CAT activity initially increased and later decreased.	Wang et al. (2012)
30.	Azinphos methyl	<i>Eisenia fetida andrei</i>	Reduced burrowing activity and inhibition of cholinesterase activity.	Jordaan et al. (2012)
31.	Dimethoate	<i>Eisenia kinneari</i>	A disturbance in the cellular enzyme system which led to profound changes in testes.	Leena et al. (2012)
32.	Fomesafen	<i>Eisenia fetida</i>	Low doses may not cause oxidative stress and peroxidation.	Zhang et al. (2013)
33.	Chlorpyrifos	<i>Eisenia fetida</i>	With increasing doses, the earthworm body began to melt and disintegrate, and thus hindering the development, growth, reproduction, and the digging process.	Pawar & Ahmad (2014)
34.	Dimethoate	<i>Eudichogaster kinneari</i>	Effects on neurosecretory cells of the brain.	Lakhani (2015)
35.	Imidacloprid	<i>Aporrectodea caliginosa</i> , <i>Lumbricus terrestris</i> , <i>Eisenia fetida</i>	Severe impacts on CNS also inhibited cellulose and SOD activity, and induced sperm deformity also found serious DNA damage.	Taillebois et al. (2018); Zhao et al. (2018); Wang et al. (2016)
36.	Chlorpyrifos, Cypermethrin	<i>Eudrilus eugeniae</i>	Variation in morpho-behavioral changes like coiling, clitellar enlargement, mucus production, bleeding, and body segmentation. Significant alterations in stress markers such as AChE, SOD, CAT, and GST.	Tiwari et al. (2019)
37.	Triazophos and Deltamethrin	<i>Eudrilus eugeniae</i>	Significant changes in oxidative stress indicators, including non-enzymatic (MDA, GSH) and enzymatic (SOD, CAT, GST, LDH, ACP, ALP, ALT, and AST).	Singh et al. (2020)
38.	Malathion	<i>Eudrilus eugeniae</i>	Effect on anatomical symptoms such as coiling, unusual swelling, mucous production, bleeding, and fragmentation was observed after 24 and 48 hours of exposure. AChE enzyme activity was inhibited.	Jeyaprakasam et al. (2021)
39.	Chlorpyrifos, Cypermethrin and Glyphosate	<i>Eudrilus eugeniae</i>	Significant increase in glutamic acid (~91.81%), lysine (~92.20%), asparagine (~94.20%), leucine (~90.20%), malic acid (~93.37%), turanose (~95.04%), Oleic acid (~93.47%), methionine (~92.27%), maltose (~92.36%), cholesta-3,5-diene (~86.11%), and galactose (~).	Malla et al. (2023)
40.	Quinalphos	<i>Eudrilus eugeniae</i>	Significant changes in stress markers. Morphological anomalies like bulging, coiling, and bleeding.	Sujeeth et al. (2023)

### Impact of pesticides on growth, development and reproduction of earthworms

The stress brought on by the presence of a contaminant, such as a pesticide, may induce the organism to redirect its energy from growth and development to survival (Gibbs et al., 1996). In order to evaluate any adverse effects of pesticides on earthworms, many scientists have considered the growth, development, and survival of earthworms as a crucial metric (Yasmin & D'Souza, 2010). Several previous works have reported the adverse effect of selected pesticides on the growth, development, and reproduction of earthworms. Mosleh et al. (2003) assessed the impact of cypermethrin, aldicarb, chlorfluazuron, profenofos, metalaxyl, and atrazine, and Booth et al. (2000) investigated the effect of two organophosphate pesticides diazinon and chlorpyrifos on

the earthworm *Aporrectodea caliginosa* and found a decrease in growth rate in the pesticide-treated worms. Another investigation by Mosleh et al. (2002, 2003) assessed the impact of endosulfan and aldicarb on the earthworm *Lumbricus terrestris* and found that the pesticides exposure has resulted into an adverse effect on the growth of the earthworms. Booth & O'Halloran (2001) and Zhou et al. (2007) separately concluded that the adverse effects of pesticides are more prominent in the juvenile stage in comparison to the adult stage. According to Zhou et al. (2006), earthworm weight was a more sensitive indicator of acetochlor and methamidophos toxicity than death. In another study, it was reported that weight loss also appears to be a valuable indicator of physiological stress, related to the degree of intoxication and time of exposure (Frampton et al., 2006).

Another symptom observed in 100% of the worms treated with Parathion was coiling, which is associated with weight loss and is thought to be the result of a change in muscle function brought on by organophosphate pesticides. This may account for the worms' difficulty moving around and relative incapacity to feed themselves (Bustos-Obregón & Goicochea, 2002). Jordaan et al. (2012) reported that the pesticide azinphos-methyl has an adverse effect on maturation, growth, reproduction, and burrowing activity and could result in cholinesterase (ChE) inhibition in *E. andrei*. According to Yasmin & D'Souza (2007), pesticides have a dose-dependent influence on worm reproduction, which may result in longer incubation times, fewer hatchlings per cocoon on average and maximum, and more worms creating cocoons overall. Several reproductive parameters have been assessed for the impact of pesticides on earthworms, like sperm maturation and viability, cocoon and hatchling production, and viability of hatchlings (Robidoux et al., 2000; De Silva et al., 2010). Several previous works, like Van Gestel (1992) and Gupta & Saxena (2003), have reported that cocoon hatchability was found to be a sensitive parameter for toxicity assessment of parathion carbendazim, pentachlorophenol, and copper oxychloride, whereas cocoon production was found to be a significant parameter for the assessment of toxic impacts of dieldrin, paraquat, fentin, benomyl, carbaryl, phenmedipham, and copper oxychloride. Bustos-Obregón & Goicochea (2002) evaluated the effects of parathion exposure on *Eisenia fetida* reproductive parameters, such as genotoxicity on male germ cells and the production of sperm and cocoons. They discovered that changes in these parameters were evident in the number of worms, sperm, and cocoons that were born. Xiao et al. (2006) reported the number of juveniles per cocoon as a sensitive parameter for assessment of the impact of acetachlor on earthworms. Dasgupta et al. (2012) reported that the toxic impact of carbaryl, chlorpyrifos, and endosulfan has resulted in a decrease in cocoon and juvenile production.

Likewise, Gupta & Saxena (2003) reported that the toxic impact of carbaryl on the earthworm *Metaphire posthuma* could produce abnormalities in the sperm head, including a wavy head, in lower concentrations, whereas at higher concentrations the sperm head becomes amorphous and the nucleus is converted into granules deposited within the region of the wavy head. Espinoza-Navarro & Bustos-Obregon (2005) reported that malathion contamination resulted in modifications in DNA of sperms, altered the cell proliferation, and decreased the sperm viability in *Eisenia fetida*. Another study by Muangphra et al. (2016) reported that chronic exposure of chlorpyrifos to the earthworm *Pheretima penguana* resulted in a reduction in cocoon production and viability. Gowri & Thangraj (2020) have assessed the impact of pesticide Monocrotophos on earthworms *Perionyx barotensis* and *Eudrilus eugeniae* and reported that the toxic impact of pesticide had resulted in the production of defective cocoons, increased mortality in the earthworms, and abnormalities in sperm, including oligopermia, necrospermia, and asthenospermia. Additionally, the growth, development, and reproduction in earthworms are crucial for their survival and ecological contributions. These processes involve a number of interrelated

parameters that need to be closely observed. Even though a lot of research has been done in this field so far, a number of different earthworm species and types still need to be examined, in contrast to a lot of various pesticide combinations. With crop yields becoming more and more dependent on agrochemicals like pesticides worldwide, there is still more study to be done in this area to better understand how the growth, development, and reproduction of the ecologically significant earthworm species interact with agrochemicals like pesticides.

### Impact of pesticides on histological parameters of earthworms

Some of the commonly used pesticides and their impact on histological parameters of earthworm species are listed in Table 2. Earthworms are extremely sensitive to changes in their surrounding environment, and therefore exposure to any chemical toxicant may produce several adverse effects, which prominently include alterations in the histological features of the earthworms. Singh et al. (2020) studied the histopathological impact of sublethal concentrations of triazophos on earthworm *E. eugenia* and reported that the earthworms exposed to the pesticides displayed both significant morphological changes in the earthworm body and histological changes in the post-clitellar region. Nunes et al. (2016) reported that the exposure of Abamectin to earthworm *Eisenia Andrei* resulted in skin thinning and discolouration, body constriction, posterior segment fragmentation, etc. In another study, Oluah et al. (2010) observed that the earthworm species *N. mbae* subjected to the pesticide atrazine had histopathological effects such as pyknotic cells, glandular growth of the epithelial tissues, and damage to the chloragogenous layer and epithelial tissues. Furthermore, Kavitha et al. (2020) reported significant histopathological changes in the intestine of *L. mauritii* after application of a lower sublethal concentration of monocrotophos; prominent damages were observed on the first few days (1<sup>st</sup> and 5th day), which thereafter slowly recovered (30th day). The intestinal tract in this investigation displayed cell fusion, pyknotic nuclei development, damaged villi, and loss of the thick epithelial lining on the first day of the experiment. On the fifth day, the cytoplasm developed vacuoles, which led to space formation, blood sinus congestion, and significantly damaged nuclei, along with extensive damage to the gut. By the thirty-first day, the epithelial cells had been regenerated and were more or less evenly spaced across the epithelial layer. Moreover, the detrimental effects of the herbicide butachlor on the earthworm species *Perionyx sansibaricus* and *Eisenia fetida* were investigated by Muthukaruppan et al. (2005) and Gobi & Gunasekaran (2010), respectively. They observed enlargement of glandular cells, visible epithelial cell damage, and cellular enlargement, along with the loss of chromatin material in the cells, demonstrating the histological as well as cytological impact of the pesticides on earthworms. Moreover, in another experiment with earthworm species *E. fetida*, the epithelium layer of the earthworm was reported to be severely damaged, resulting in fused cells and excessive villous development. There were necrotic cell ruptures, causing widespread dispersion of cell debris. The chloragogen tissue was also

severely damaged with minimal reserve inclusion. It was also observed that with the increase in the dose concentration of the pesticide, the villi continued to get fused while a distinct cavitation and pyknotic nuclei were observed in the epithelial layer of the gut of the earthworm. Also, the previous studies have clearly demarcated the histological alterations and damage caused by pesticide exposure. However, further research is required to fully comprehend how histological changes are related to the type and dosage of pesticides that are commonly used, especially in agricultural fields, as new types of chemical combinations of pesticides are regularly emerging for crop protection. The histological impacts of any pesticide may be assessed in different tissue types, or the comparative histological impacts may be analyzed on a single tissue type following the exposure to different pesticide combinations. Further, the molecular mechanisms and gene expression analysis may be done to comprehend how a pesticide is triggering the molecular mechanisms leading to necrosis or cell death.

### Impact of pesticides on regeneration of earthworms

Earthworms are soil-dwelling animals; hence, they are very much prone to serious damage and wounds inflicted on them, and therefore they have a great regenerative capacity, which helps in their survival. Numerous reports have demonstrated the adverse effects of pesticides on earthworms, but only a few works have been conducted to study the pernicious effects of pesticides on the regenerative capacity of earthworms. According to an experiment conducted by Kumari & Sinha (2011), different

doses of malathion on all the segments at different intervals significantly ( $p < 0.01$ ) affected the process of regeneration in the earthworm *D. willsi*, a cropland earthworm. In another experiment, Zoran et al. (1986) observed the teratogenic effects of benomyl on the earthworm species *Eisenia fetida* and reported that there was a reduction in the number of regenerated segments and an increased frequency of segmental groove anomalies, followed by a variety of other abnormalities. The effects of the pesticide benomyl treatment on the number of regenerated segments and frequency of anomalies were found to be dose-dependent within a narrow concentration range (approximately 0.2–5.0 mg/litre) and at higher concentrations (e.g., 25 mg/litre); teratogenic effects were less frequent because of the delayed onset of segmental delineation until after exposure. Furthermore, Yuan et al. (2024) evaluated the impact of Carbamates, Metolcarb, and Fenoxycarb on *Eisenia fetida* and reported that though the precise mechanisms of harmful effects of CAR's (Carbamates) on earthworms remain unclear, especially from a regenerative standpoint, MEB (Metolcarb) and FEB (Fenoxycarb) treatments resulted in delayed posterior renewal of amputated earthworms compared to the control group, resulting in compromised morphology, dwindling segments, and increased cell apoptosis in blastemal tissues. This was mediated by rising Sox2 (sex-determining region Y-box 2) and decreasing TCTP (translationally controlled tumor protein) levels. These findings provide baseline toxicity cues for MEB and FEB exposure against earthworms, as well as mechanistic insights into regenerative toxicity during CAR exposure. Previous studies in

**Table 2.** Impact of some commonly used Pesticides on histological parameters of earthworms.

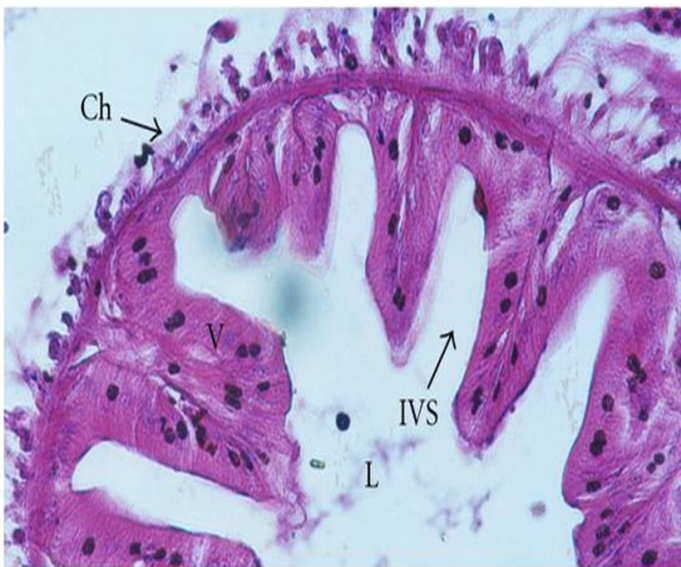
S.No.	Pesticide	Earthworm species	Effect	References
1.	Benomyl	<i>Eisenia fetida</i>	Hypodeveloped sperms; sperms with abnormal acrosome development.	Sorour & Larink (2001)
2.	Butachlor	<i>Perionyx sansibaricus</i>	Glandular cell enlargement at all exposed concentrations.	Gobi et al. (2005)
3.	Malathion	<i>Eisenia fetida</i>	Decreased spermatic viability in spermatheca, altering the cell proliferation and modifying the DNA structure of spermatogonia.	Espinoza-Navarro & Bustos-Obregón (2005)
4.	Carbofuran	<i>Eisenia fetida</i>	Protein content and total cholinesterase activity (TChE) increased in low level.	Ling (2006)
5.	Profenofos	<i>Eisenia fetida</i>	Body ruptures and bloody lesions.	Reddy & Rao (2008)
6.	Butachlor	<i>Eisenia fetida</i>	The epithelium layer of the earthworm was reported to be severely damaged, resulting in fused cells and excessive villous development.	Gobi et al. (2010)
7.	Atrazine	<i>Nsukkadrilus mbae</i>	Chloragogenous layer and epithelial tissue damage; prominent vacuolations and pyknotic cells.	Oluah et al. (2010)
8.	Triplen, Ridomil, Cyren and Mamba	<i>Lumbricus terrestris</i>	Symptoms of toxicity found were body swellings, coelomicfluid discharge, slow movements, coiling etc..	Ahmed (2013)
9.	Imidacloprid, Acetamiprid, Nitenpyram, Clothianidin and Thiacloprid	<i>Eisenia fetida</i>	Significantly inhibit fecundity and cellulase activity and also damage the epidermal and midgut cells.	Wang et al. (2015)
10.	Abamectin	<i>Eisenia andrei</i>	Skin thinning and discoloration, body constriction and posterior segment fragmentation.	Nunes et al. (2016)
10.	Monocrotophos	<i>Lampito mauritii</i>	Intestinal tract showed loss of dense epithelial lining, damaged villi, cell fusion, and formation of pyknotic nuclei.	Kavitha et al. (2020)
11.	Monocrotophos	<i>Eudrilus eugeniae</i> and <i>Perionyx barotensis</i>	Increase in mortality and abnormal sperm (asthenospermia, necrospermia, and oligospermia). Rupture of chloragogenous tissue, longitudinal muscle, fused and extra-villous growth and necrotic cell rupture in earthworm's body wall (epidermis, circular and longitudinal muscles) were observed.	Gowri & Thangaraj (2020)
12.	Chlorpyrifos	<i>Eudrilus euginae</i>	The gut region showed mild degeneration of intestinal epithelium, epithelial cells, and fusion of villi. Epidermal hyperplasia was also observed.	Krishnaswamy et al. (2021)

molecular mechanisms involved in regeneration reveal that major proteinases, like Plasmin, Plasminogen activator, and Elastases play a crucial role in spatiotemporal distribution and hydrolysis of extracellular matrix components during regeneration (Sun et al., 2001). The expression of Sox 2 protein and translationally controlled tumour proteins (TCTPs) rises significantly during regeneration (Subramaniam et al. 2017; Jing Tao et al. 2018). Other significantly involved proteins in the regeneration process include Wnt 3a (stem cell marker), PCNA (cell proliferation), and YAP 1 (hippo signalling), which are expressed in response to THTPs (Rajgopalan et al. 2022). The regeneration mechanism includes a group of intricately related mechanisms at molecular, cellular, and tissue levels, and therefore a deep under-

standing of how a pesticide affects the process of regeneration is very essential. But, still, not much work has been done on this aspect, and hence further studies are required to comprehend how various pesticides affect the physiological, anatomical, or molecular processes involved in earthworm regeneration.

### Impact of pesticides on biochemical and physiological parameters of earthworms

Some of the commonly used pesticides and their impact on biochemical parameters of earthworm species are listed in Table 3. Through the course of various studies and research, a variety of remarkable observations were made regarding the impact of pesticides on different biochemical and physiological parameters of earthworms. According to Datta et al. (2016), earthworms constitute up to 60–80% biomass in the soil macrofauna and are prone to the non-target effects of soil pesticide contamination as it's widely distributed in agroecosystems. Almost all organophosphate pesticides are neurotoxic, and generally, upon acute exposure, acetylcholinesterase (AChE) enzyme activity is inhibited, resulting in the accumulation of acetylcholine at nerve and muscle endings, which stimulates both muscarinic and nicotinic receptors in the nerves and muscles (Thompson & Richardson, 2004). The exposure of CARs like carbofuran (Ferreira et al., 2015; Kanedi, 2017a, 2017b), carbaryl (Lima et al., 2015), aldicarb (Mosleh et al., 2003), and methomyl (Ibtissem et al., 2012) results in negative physiological responses of earthworms, which involve avoidance behaviour, decreased reproduction, weight loss, reduced survival rates, and impaired enzymatic system. According to Wang et al. (2015), the DNA in the cell nucleus, which was supposedly exposed to the highest concentration (1.00 mg/kg imidacloprid), was fluffy, scattered, and dusky, with the DNA migrating into the tail region as a result of extensive strand breakage, and as the concentrations decreased, the DNA migrating extended in the cell nucleus. It was also seen that the nuclear DNA migrated the most significantly from the cell nuclear space after it was exposed to the lowest concentration (0.10 mg/kg imidacloprid). In another work carried out by Qiao et al. (2022), it was observed that the subchronic toxicity of FLU (Flupyradifurone) resulted in an increase in the activities of superoxide dismutase (SOD), catalase (CAT), and glutathione-S transferase (GST), while the peroxidase (POD) and acetylcholinesterase (AChE) activities decreased. In this study, it was also revealed that the reactive oxygen species (ROS) level and malondialdehyde (MDA) content in earthworms were increased by FLU, resulting in DNA damage. Based on several scientific reports, it has been established that the pesticides may produce oxidative stress, which might result in severe adverse effects like DNA damage and abnormal levels of antioxidant enzymes in the body, among others, which can adversely affect growth, survival, and other facets of earthworm biology. Therefore, more work is needed in this area to understand how different free radicals are generated as a result of exposure to different pesticides and how they damage or hamper vital molecular and physiological mechanisms within the cells.



**Figure 1.** Cross section of earthworm *E. fetida* normal intestine and chloragogen tissue at control. L: Lumen; V: Villi; Ch: chloragogen tissue; IVS: Inter villous space (Courtesy: Gobi & Gunasekaran, 2010).



**Figure 2.** Cross section of earthworm *E. fetida* showing intestine and chloragogen tissue at 0.2575 mg kg<sup>-1</sup> concentration of herbicide. PK: pyknotic nuclei; CD: Cellular Debris; L: Lumen; FV: Fused Villi; Ch: chloragogen tissue (Courtesy: Gobi & Gunasekaran, 2010).

**Table 3.** Impact of pesticides on biochemical parameters of earthworms.

S. No.	Pesticide	Earthworm species	Enzymes	Effect	References
1.	Azoxystrobin	<i>Eisenia fetida</i>	SOD and GST	Both the SOD and GST activities were increased.	Han et al. (2014)
2.	Imidacloprid	<i>Eisenia fetida</i>	SOD	SOD activity increased at lower dose but decreased at higher dose.	Zhang et al. (2014)
3.	Imidacloprid	<i>Eisenia fetida</i>	AChE	AChE activity was suppressed.	Wang et al. (2015)
4.	Carbaryl	<i>Eudrilus eugenie</i> , <i>Perionyx ceylanensis</i> and <i>Perionyx excavatus</i>	SOD, CAT, GST	SOD and CAT activity was suppressed whereas there was an increase in GST.	Jeyanthi et al. (2016)
5.	Carbendazim and Captan	<i>Eisenia fetida</i>	ACP	ACP activity was inhibited.	Mandal et al. (2017)
6.	Clothianidin	<i>Eisenia fetida</i>	GST	GST activity was decreased at higher concentration.	Liu et al. (2017)
7.	Glyphosate	<i>Dendrobaena veneta</i>	AChE	AChE activity was significantly increased.	Hackenberger et al. (2018)
8.	Fluoxastrobin	<i>Eisenia fetida</i>	SOD and CAT	SOD activity was increased, whereas CAT activity was suppressed.	Zhang et al. (2018)
9.	Chlorpyrifos and Cypermethrin	<i>Eudrilus eugeniae</i>	AChE	Inhibition of AChE activity was region and dose-dependent.	Tiwari et al. (2019)
10.	Pyraclostrobin	<i>Eisenia fetida</i>	SOD	SOD activity was increased in beginning and reduced with time.	Ma et al. (2019)
11.	Sulfentrazone	<i>Eisenia fetida</i>	SOD, CAT, GST	SOD was greatly suppressed at the beginning but steadily enhanced with increasing concentration. GST activity was higher in comparison to control among all the three enzymes.	Li et al. (2020)
12.	Triazophos and Deltamethrin	<i>Eudrilus eugeniae</i>	ACP and ALP	Both ACP and ALP activities were significantly increased.	Singh et al. (2020)
13.	Tebuconazole	<i>Eisenia fetida</i>	CAT	No changes were reported.	Zhang et al. (2020)
14.	Sulfoxafor	<i>Eisenia fetida</i>	GST	GST activity was enhanced.	Zhang et al. (2020)
15.	Glyphosate	<i>Alma millsoni</i> , <i>Eudrilus eugeniae</i> and <i>Libyodrilus violaceus</i>	AChE	No significant changes were observed.	Owagboriaye et al. (2020)
16.	Trifloxystrobin	<i>Eisenia fetida</i>	SOD	SOD activity did not vary much at low concentrations but improved considerably at higher dose.	Liu et al. (2020)
17.	Chlorpyrifos	<i>Eisenia fetida</i>	CAT	CAT activity was severely inhibited at higher concentrations, whereas there was no significant difference seen at low concentrations.	Zhu et al. (2020)
18.	Thifuzamide	<i>Eisenia fetida</i>	SOD and GST	SOD activity was inhibited at higher doses whereas GST activity was dose dependent.	Yao et al. (2020)
19.	Tebuconazole	<i>Eisenia fetida</i>	CAT and SOD	SOD activity considerably decreased and CAT activity showed no effect.	Zhang et al. (2020)
20.	Atrazine	<i>Eisenia fetida</i>	ALP	ALP activity was decreased.	Lammertyn et al. (2021)
21.	Trifloxystrobin	<i>Eisenia fetida</i>	CAT	CAT activity was decreased.	Wu et al. (2021)

### Impact of pesticides on gut microbiota of earthworms

The unique features of the gut ecosystem in earthworms play an important role in improving the soil fertility parameters during the gut transition of soil. It contains an array of enzymes produced endogenously by the residing microbes, used for the digestion of organic matter (Drake & Horn, 2007). The gut microbiota of earthworms contains bacteria, fungi, protozoa, and other microbes that play an important part in nutrient metabolism, digestion, and even immune functions by contributing to organic matter breakdown, nutrient release, and maintaining gut homeostasis (Furlong et al., 2002). Depending on the ecological trophic group, earthworms can contribute to further movement of pollutants into the soil (Kuzyakov & Blagodatskaya, 2015). For instance, *Lumbricus terrestris*, an anecic earthworm (Bouché, 1977; Lavelle et al., 1989), is capable of transporting pesticides from the surface to the mineral horizons. Moreover, *L. terrestris* digestion can take up to 6 hours (Nechitaylo et al., 2010) due to the structural features like a longer intestinal tract; consequent-

ly, when pesticides enter this digestive tract, they can have long-term impacts on gut microorganisms. According to Astaykina et al. (2022), pesticide-contaminated soil greatly affects the microbial community that lives in the guts of earthworms. They found that pesticides, imidacloprid, benomyl, and metribuzin, reduced the total bacterial diversity in the earthworm's gut even at the recommended application rate. Under the administered pesticides, the structure of the gut prokaryotic community experienced alterations in the relative abundance of the phyla proteobacteria, actinobacteria, acidobacteria, planctomyces, verrucomicrobia, and cyanobacteria. Furthermore, pesticides influenced the amount of *Verminephrobacter*—the earthworms' nephridia-specific symbionts. These findings demonstrated that the earthworm's gut microbial population is extremely vulnerable to pesticide contamination in soil. Krishnaswamy et al. (2021) studied the impact of pesticide Chlorpyrifos (CPF) toxicity on the gut microbiome of *E. euginae* by using a long-amplicon Nanopore sequencing method. The results showed no fluctua-



tions with Firmicutes and Bacteroidetes, which were shown to be dominant at the bacterial phylum level, while significant differences were observed at the genus level. Chang *et al.* (2021) stated that the pesticide Fomesafen not only targeted and significantly altered the gut microbial phyla but also that exposure to this biocide lowered the earthworms' energy resources and activated the antioxidant system, both of which were strongly linked with gut microbial diversity. Similarly, three species of earthworms, namely *Alma millsoni*, *Eudrilus eugeniae*, and *Libyodrilus violaceus*, when exposed to a glyphosate-based herbicide, showed a significant shift in bacterial populations, with Proteobacteria becoming the dominant phylum; the affected bacterial genera mostly belonged to genera *Pantoea*, *Pseudomonas*, and *Enterobacter* (Owagboriaye *et al.*, 2021). Moreover, Kavitha *et al.* (2020) worked on the impact of monocrotophos on *Lampito mauritii*, an Indian earthworm species commonly known as Kinberg, and observed that after 30 days of exposure, microbial populations returned to their normal levels after an initial decrease in microbiota. This work revealed that only six bacterial and three fungal species survived exposure to monocrotophos out of eight bacterial and five fungal species. On the first and fifth days of the experiment, the gut showed severe pathological alterations, including vacuolization, degraded nuclei, damaged villi, and blood sinus congestion. But on the 30th day, the damages were slowly recovered due to the degradation of monocrotophos by the presence of some pesticide-degrading bacterial and fungal species and the regenerative capability of chloragogen cells in the intestine. Prior research has demonstrated that interactions between gut microbiota and the transient soil in the gut of earthworms have a significant role in improving soil fertility. Any external factor, such as pesticides, that can adversely affect the microbial environment or microbial diversity of the earthworm gut will hamper soil processing through the earthworm gut, consequently affecting the biology and ecological role of earthworms. Further, the microbial diversity of earthworm gut has been found to play a role in mitigation of pesticide contamination, and therefore more studies in this area are required for a better understanding of the interaction between gut microbiota and the chemical contaminants like pesticide exposure to the earthworms. Despite earlier studies in this area, additional research is necessary to fully understand how different pesticides can alter earthworm gut microbiota and what impact this interaction may have on the role of earthworms in soil ecosystem dynamics.

## Conclusion

Even though pesticides tend to play a significant role in better crop yield, it also significantly destroys and handicaps the natural biological soil fertility creatures which include the earthworms. Studies consistently demonstrate adverse effects of pesticides in earthworms, such as reduced population, altered behavior, histopathological alterations, impaired growth, development and reproduction, altered regenerative capacities and adverse changes in gut microbiota among other marked effects. It is quite evident that there are numerous chemical combinations that are used as

pesticides and on the other hand there are different ecological groups and species of earthworms. A thorough understanding of different aspects of the effects of pesticides on earthworm biology, from the molecular, cytological, histological, physiological, individual, and ecological levels, is still lacking despite a number of earlier studies. Any factor that can adversely affect the earthworm biology can have cascading effects on other soil organisms, plant communities, soil structure, and ultimately, ecosystem health. These findings underscore the need of further research in various aspects of earthworm-pesticides interactions which will prove to be significant for sustainable agricultural practices to mitigate harm to earthworm populations and maintain soil biodiversity, fertility and soil ecosystem dynamics.

## DECLARATIONS

### Author contribution statement

Conceptualization: A.K.; Methodology: A.K. and N.S.; Software and validation: N.S. and K.A.; Formal analysis and investigation: A.K. and N.S.; Resources: N.S. and S.F.; Data curation: N.S., K.A. and S.F.; Writing-original draft preparation: N.S., K.A. and S.F.; Writing-review and editing: A.K.; Visualization: K.A., S.F. and N.S.; Supervision: A.K.; Project administration: A.K.; Funding acquisition: A.K.; All authors have read and agreed to the published version of the manuscript.

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## REFERENCES

- Abdul, R. A., & Bouche, M. B. (1997). Earthworm toxicology: From acute to chronic tests. *Soil Biology and Biochemistry*, 29(3-4), 699-703.
- Ahmad, L., Khan, A., Khan, M. Z., Hussain, I., Mahmood, F., Sleemi, M. K., Lodhi, L. A., & Abdullah, I. (2012). Toxicopathological effects of cypermethrin upon male reproductive system in rabbits. *Pesticide Biochemistry and Physiology*, 103(3), 194-201.
- Ahmed, S. T. (2013). The impact of four pesticides on the earthworm *Lumbricus terrestris* (Annelida; Oligochaeta). *International Journal of Current Research and Review*, 5(21), 1.
- Aina, P. O. (1984). Contribution of earthworms to porosity and water infiltration in a tropical soil under forest and long-term cultivation. *Pedobiologia*.
- Alavanja, M. C. (2009). Introduction: Pesticides use and exposure, extensive worldwide. *Reviews on Environmental Health*, 24(4), 303-310.
- Amenyogbe, E., Huang, J. S., Chen, G., & Wang, Z. (2021). An overview of the pesticides' impacts on fishes and humans. *International Journal of Aquatic Biology*, 9(1), 55-65.
- Anjum, M. M., Ali, N., & Iqbal, S. (2017). Pesticides and environmental health: A review. *Toxicology and Environmental Chemistry*, 5, 555671.
- Astaykina, A., Streletskii, R., Maslov, M., Krasnov, G., & Gorbato, V. (2022). Effects of three pesticides on the earthworm *Lumbricus terrestris* gut microbiota. *Frontiers in Microbiology*, 13, 853535.
- Bardiya, S., & Dixit, S. (2024). A review on effect of different pesticides on earthworm. *International Journal of Science and Research Archive*, 11(1), 590-599.
- Bhadauria, T., & Saxena, K. G. (2010). Role of earthworms in soil fertility maintenance through the production of biogenic structures. *Applied and Environmental Soil Science*, 2010(1), 816073.
- Bond, J. L., Kriesemer, S. K., Emborg, J. E., & Chadha, M. L. (2009). Understanding farmers' pesticide use in Jharkhand India. *Extension Farming Systems Journal*, 5(1), 53-61.
- Booth, L. H., & O'Halloran, K. (2001). A comparison of biomarker responses in the earthworm *Aporrectodea caliginosa* to the organophosphorus insecticides diazinon and chlorpyrifos. *Environmental Toxicology and Chemistry: An International Journal*, 20(11), 2494-2502.
- Booth, L. H., Heppelthwaite, V. J., & O'halloran, K. (2000). Growth development and fecundity of the earthworm *Aporrectodea caliginosa* after exposure to two organophosphates. *New Zealand Plant Protection*, 53, 221-225.
- Bouché, M. B. (1977). Strategies lombriciennes. *Ecological Bulletins*, 122-132.
- Brown, G. G., Benito, N. P., Pasini, A., Sautter, K. D., de F Guimarães, M., & Torres, E. (2003). No-tillage greatly increases earthworm populations in Paraná state, Brazil: The 7th international symposium on earthworm ecology. Cardiff, Wales. 2002. *Pedobiologia*, 47(5-6), 764-771.
- Buchel, K. H. (Ed.). (1983). *Chemistry of pesticides* (pp. xii+518).
- Bustos-Obregón, E., & Goicochea, R. I. (2002). Pesticide soil contamination mainly affects earthworm male reproductive parameters. *Asian Journal of Andrology*, 4(3), 195-199.
- Capowiez, Y., & Bérard, A. (2006). Assessment of the effects of imidacloprid on the behavior of two earthworm species (*Aporrectodea nocturna* and *Allolobophora icterica*) using 2D terraria. *Ecotoxicology and Environmental Safety*, 64(2), 198-206.
- Capowiez, Y., Rault, M., Costagliola, G., & Mazzia, C. (2005). Lethal and sublethal effects of imidacloprid on two earthworm species (*Aporrectodea nocturna* and *Allolobophora icterica*). *Biology and Fertility of Soils*, 41(3), 135-143.
- Casabé, N., Piola, L., Fuchs, J., Oneto, M. L., Pamparato, L., Basack, S., Giménez, R., Massaro, R., Papa, J. C., & Kesten, E. (2007). Ecotoxicological assessment of the effects of glyphosate and chlorpyrifos in an Argentine soya field. *Journal of Soils and Sediments*, 7, 232-239.
- Chang, X., Sun, Y., Zhao, L., Li, X., Yang, S., Weng, L., & Li, Y. (2021). Exposure to fomesafen alters the gut microbiota and the physiology of the earthworm *Pheretima guillelmi*. *Chemosphere*, 284, 131290.
- Cho, S. J., Cho, P. Y., Lee, M. S., Na, Y., Lee, J. H., Koh, K. S., Choo, J.K., & Park, S. C. (2001). Up-regulation of multiple serine proteinases during earthworm tail regeneration. *Invertebrate Reproduction & Development*, 40(2-3), 103-108.
- DasGupta, R., Chakravorty, P. P., & Kaviraj, A. (2012). Effects of carbaryl, chlorpyrifos and endosulfan on growth, reproduction and respiration of tropical epigeic earthworm, *Perionyx excavatus* (Perrier). *Journal of Environmental Science and Health, Part B*, 47(2), 99-103.
- Datta, S., Singh, J., Singh, S., & Singh, J. (2016). Earthworms, pesticides and sustainable agriculture: a review. *Environmental Science and Pollution Research*, 23(9), 8227-8243.
- De Silva, P. M. C., Pathiratne, A., & van Gestel, C. A. (2010). Toxicity of chlorpyrifos, carbofuran, mancozeb and their formulations to the tropical earthworm *Perionyx excavatus*. *Applied Soil Ecology*, 44(1), 56-60.
- Dittbrenner, N., Moser, I., Triebkorn, R., & Capowiez, Y. (2011). Assessment of short and long-term effects of imidacloprid on the burrowing behaviour of two earthworm species (*Aporrectodea caliginosa* and *Lumbricus terrestris*) by using 2D and 3D post-exposure techniques. *Chemosphere*, 84(10), 1349-1355.
- Dittbrenner, N., Triebkorn, R., Moser, I., & Capowiez, Y. (2010). Physiological and behavioural effects of imidacloprid on two ecologically relevant earthworm species (*Lumbricus terrestris* and *Aporrectodea caliginosa*). *Ecotoxicology*, 19, 1567-1573.
- Drake, H. L., & Horn, M. A. (2007). As the worm turns: the earthworm gut as a transient habitat for soil microbial biomes. *Annual Review in Microbiology*, 61(1), 169-189.
- Edwards, C. A., & Bohlen, P. J. (1996). *Biology and ecology of earthworms* (Vol. 3). Springer Science & Business Media.
- Espinoza-Navarro, O., & Bustos-Obregón, E. (2005). Effect of malathion on the male reproductive organs of earthworms, *Eisenia foetida*. *Asian Journal of Andrology*, 7(1), 97-101.
- Faheem, M., & Khan, M. F. (2010). Toxicity of imidacloprid (Nicotinoid) against earthworm, *Pheretima posthuma* with reference to its effects on protein. *Journal of Basic and Applied Science*, 6(1), 55-62.
- Ferreira, R. C., Papini, S., & de Andréa, M. M. (2015). Bioavailability and influence of 14C-carbofuran on *Eisenia andrei* avoidance, growth and reproduction in treated natural tropical soils. *Journal of Environmental Science and Health, Part B*, 50(4), 266-274.
- Frampton, G. K., Jänsch, S., Scott-Fordsmann, J. J., Römbke, J., & Van den Brink, P. J. (2006). Effects of pesticides on soil invertebrates in laboratory studies: a review and analysis using species sensitivity distributions. *Environmental Toxicology and Chemistry: An International Journal*, 25(9), 2480-2489.
- Furlong, M. A., Singleton, D. R., Coleman, D. C., & Whitman, W. B. (2002). Molecular and culture-based analyses of prokaryotic communities from an agricultural soil and the burrows and casts of the earthworm *Lumbricus rubellus*. *Applied and Environmental Microbiology*, 68(3), 1265-1279.
- Gambi, N., Pasteris, A., & Fabbri, E. (2007). Acetylcholinesterase activity in the earthworm *Eisenia andrei* at different conditions of carbaryl exposure. *Comparative Biochemistry and Physiology Part C: Toxicology & Pharmacology*, 145(4), 678-685.
- Gibbs, M. H., Wicker, L. F., & Stewart, A. J. (1996). A method for assessing sublethal effects of contaminants in soils to the earthworm, *Eisenia foetida*. *Environmental Toxicology and Chemistry: An International Journal*, 15(3), 360-368.
- Gobi, M., & Gunasekaran, P. (2010). Effect of butachlor herbicide on earthworm *Eisenia fetida*—its histological perspicuity. *Applied and Environmental Soil Science*, 2010(1), 850758.
- Gowri, S., & Thangaraj, R. (2020). Studies on the toxic effects of agrochemical pesticide (Monocrotophos) on physiological and reproductive behavior of indigenous and exotic earthworm species. *International Journal of Environmental Health Research*, 30(2), 212-225.
- Greig-Smith, P. W. (1992). *Ecotoxicology of Earthworms*. Intercept.
- Gupta, S. K., & Saxena, P. N. (2003). Carbaryl-induced behavioural and reproductive abnormalities in the earthworm *Metaphire posthuma*: a sensitive model. *Alternatives to Laboratory Animals*, 31(6), 587-593.
- Hackenberger, D. K., Stjepanović, N., Lončarić, Ž., & Hackenberger, B. K. (2018). Acute and subchronic effects of three herbicides on biomarkers and reproduction in earthworm *Dendrobaena veneta*. *Chemosphere*, 208, 722-730.
- Han, Y., Zhu, L., Wang, J., Wang, J., Xie, H., & Zhang, S. (2014). Integrated assessment of oxidative stress and DNA damage in earthworms (*Eisenia fetida*) exposed to azoxystrobin. *Ecotoxicology and Environmental Safety*, 107, 214-219.
- Hole, D. G., Perkins, A. J., Wilson, J. D., Alexander, I. H., Grice, P. V., & Evans, A. D. (2005). Does organic farming benefit biodiversity? *Biological Conservation*, 122(1), 113-130.
- Ibtissem, Z., Aicha, T., Nedjoud, G., Nawel, B., Houria, B., & Reda, D. M. (2012). Potential toxicity of an insecticide of the family of carbamates on a bioindicator model of the pollution the earthworm *Octodrilus complanatus* (Oligochaeta, Lumbricidae). *Annals of Biological Research*, 3(11), 5367-5373.
- Jeyanthi, V., Paul, J. A. J., Selvi, B. K., & Karmegam, N. (2016). Comparative study of biochemical responses in three species of earthworms exposed to pesticide and metal contaminated soil. *Environmental Processes*, 3, 167-178.

- Jeyaprakasam, A., Muniyandi, B., James, A. J. P., Karmegam, N., & Ponnuchamy, K. (2021). Assessment of earthworm diversity and pesticide toxicity in *Eudrilus eugeniae*. *Environmental Chemistry and Ecotoxicology*, 3, 23-30.
- Jordaan, M. S., Reinecke, S. A., & Reinecke, A. J. (2012). Acute and sublethal effects of sequential exposure to the pesticide azinphos-methyl on juvenile earthworms (*Eisenia andrei*). *Ecotoxicology*, 21, 649-661.
- Kanedi, M. (2017). Inhibitory Effect of Carbofuran Furadan 3G on the Cocoon Production and Viability in *Pheretima javanica* Gates. *IOSR Journal of Environmental Science, Toxicology and Food Technology*, 11(5 Vers), 59-62.
- Kanedi, M. (2017). Physiological effects of carbofuran on earthworm *Pheretima javanica* Gates. *Advances in Life Sciences*, 7(2), 21-25.
- Kavitha, V., Anandhan, R., Alharbi, N. S., Kadaikunnan, S., Khaled, J. M., Almanaa, T. N., & Govindarajan, M. (2020). Impact of pesticide monocrotophos on microbial populations and histology of intestine in the Indian earthworm *Lampito mauritii* (Kinberg). *Microbial Pathogenesis*, 139, 103893.
- Krishnaswamy, V. G., Jaffar, M. F., Sridharan, R., Ganesh, S., Kalidas, S., Palanisamy, V., & Mani, K. (2021). Effect of chlorpyrifos on the earthworm *Eudrilus eugeniae* and their gut microbiome by toxicological and metagenomic analysis. *World Journal of Microbiology and Biotechnology*, 37, 1-12.
- Kumari, T., & Sinha, M. (2011). Effects of sublethal doses of Malthion on regeneration of earthworm *D. willsi*. *The Ecoscan*, 155-9.
- Kuz'yakov, Y., & Blagodatskaya, E. (2015). Microbial hotspots and hot moments in soil: concept & review. *Soil Biology and Biochemistry*, 83, 184-199.
- Lakhani, L. (2015). Impact of Dimethoate on brain of the earthworm *Eudichogaster kinneari* (Stephenson): A histological profile. *Environment Conservation Journal*, 16(1&2), 1-8.
- Lammertyn, S., Masin, C. E., Zalazar, C. S., & Fernandez, M. E. (2021). Biomarkers response and population biological parameters in the earthworm *Eisenia fetida* after short term exposure to atrazine herbicide. *Ecological Indicators*, 121, 107173.
- Lavelle, P., Barois, I., Martin, A., Zaidi, Z., & Schaefer, R. (1989). Management of earthworm populations in agro-ecosystems: a possible way to maintain soil quality? In *Ecology of Arable Land—Perspectives and Challenges: Proceeding of an International Symposium, 9-12 June 1987 Swedish University of Agricultural Sciences, Uppsala, Sweden* (pp. 109-122). Springer Netherlands.
- Leena, L., Amrita, K., & Preeti, C. (2012). Effect of dimethoate on testicular histomorphology of the earthworm *Eudichogaster Kinneari* (Stephenson). *International Research Journal of Biological Sciences*, 1(4).
- Li, M., Ma, X., Saleem, M., Wang, X., Sun, L., Yang, Y., & Zhang, Q. (2020). Biochemical response, histopathological change and DNA damage in earthworm (*Eisenia fetida*) exposed to sulfentrazone herbicide. *Ecological Indicators*, 115, 106465.
- Lima, M. P., Cardoso, D. N., Soares, A. M., & Loureiro, S. (2015). Carbaryl toxicity prediction to soil organisms under high and low temperature regimes. *Ecotoxicology and Environmental Safety*, 114, 263-272.
- Ling, H. (2006). Effect of Carbofuran on Protein Content and the SOD and TChE Activity of the *Eisenia fetida* Earthworm. *Journal of Anhui Agricultural Science*, 34, 3165.
- Lister, L. J., Svendsen, C., Wright, J., Hooper, H. L., & Spurgeon, D. J. (2011). Modelling the joint effects of a metal and a pesticide on reproduction and toxicokinetics in Lumbricid earthworms. *Environment International*, 37(4), 663-670.
- Liu, T., Liu, Y., Fang, K., Zhang, X., & Wang, X. (2020). Transcriptome, bioaccumulation and toxicity analyses of earthworms (*Eisenia fetida*) affected by trifloxystrobin and trifloxystrobin acid. *Environmental Pollution*, 265, 115100.
- Liu, T., Wang, X., Chen, D., Li, Y., & Wang, F. (2018). Growth, reproduction and biochemical toxicity of chlorantraniliprole in soil on earthworms (*Eisenia fetida*). *Ecotoxicology and Environmental Safety*, 150, 18-25.
- Liu, T., Wang, X., You, X., Chen, D., Li, Y., & Wang, F. (2017). Oxidative stress and gene expression of earthworm (*Eisenia fetida*) to clothianidin. *Ecotoxicology and Environmental Safety*, 142, 489-496.
- Lu, J., Wu, Q., Yang, Q., Li, G., Wang, R., Liu, Y., Duan, C., Duan, S., He, X., Huang, Z., and Peng, X., Yan, W., & Jiang, J. (2021). Molecular mechanism of reproductive toxicity induced by beta-cypermethrin in zebrafish. *Comparative Biochemistry and Physiology Part C: Toxicology & Pharmacology*, 239, 108894.
- Lydy, M. J., & Linck, S. L. (2003). Assessing the impact of triazine herbicides on organophosphate insecticide toxicity to the earthworm *Eisenia fetida*. *Archives of Environmental Contamination and Toxicology*, 45, 343-349.
- Ma, J., Cheng, C., Du, Z., Li, B., Wang, J., Wang, J., Wang, Z., & Zhu, L. (2019). Toxicological effects of pyraclostrobin on the antioxidant defense system and DNA damage in earthworms (*Eisenia fetida*). *Ecological Indicators*, 101, 111-116.
- Mahmood, I., Imadi, S. R., Shazadi, K., Gul, A., & Hakeem, K. R. (2016). Effects of pesticides on environment. *Plant, Soil and Microbes: Volume 1: Implications in Crop Science*, 253-269.
- Malla, M. A., Dubey, A., Kori, R. K., Sharma, V., Kumar, A., Yadav, S., & Kumari, S. (2023). GC-MS based untargeted metabolomics reveals the metabolic response of earthworm (*Eudrilus eugeniae*) after chronic combinatorial exposure to three different pesticides. *Scientific Reports*, 13(1), 8583.
- Mandal, S., Chakravorty, P. P., & Kundu, J. K. (2017). Relative toxicity of two selected fungicides on acid phosphatase and alkaline phosphatase activity of epigeic earthworm *Eisenia Fetida* (Oligochaeta). *World Wide J Multidis Res Develop/WWJMRD*, 4, 14-17.
- Mosleh, Y. Y., Ismail, S. M., Ahmed, M. T., & Ahmed, Y. M. (2003). Comparative toxicity and biochemical responses of certain pesticides to the mature earthworm *Aporrectodea caliginosa* under laboratory conditions. *Environmental Toxicology: An International Journal*, 18(5), 338-346.
- Mosleh, Y. Y., Paris-Palacios, S., Couderchet, M., & Vernet, G. (2002). Biological effects of two insecticides on earthworms (*Lumbricus terrestris* L.) under laboratory conditions. *Mededelingen (Rijksuniversiteit te Gent. Fakulteit van de Landbouwkundige en Toegepaste Biologische Wetenschappen)*, 67(2), 59-68.
- Mosleh, Y. Y., Paris, Palacios, S., Couderchet, M., & Vernet, G. (2003). Acute and sublethal effects of two insecticides on earthworms (*Lumbricus terrestris* L.) under laboratory conditions. *Environmental Toxicology: An International Journal*, 18(1), 1-8.
- Muangphra, P., Tharapoom, K., Euawong, N., Namchote, S., & Gooneratne, R. (2016). Chronic toxicity of commercial chlorpyrifos to earthworm *Pheretima peguana*. *Environmental Toxicology*, 31(11), 1450-1459.
- Muthukaruppan, G., Janardhanan, S., & Vijayalakshmi, G. (2005). Sublethal Toxicity of the Herbicide Butachlor on the Earthworm *Perionyx sansibaricus* and its Histological Changes (5 pp). *Journal of Soils and Sediments*, 5, 82-86.
- Nechitaylo, T. Y., Yakimov, M. M., Godinho, M., Timmis, K. N., Belogolova, E., Byzov, B. A., Kurakov, A. V., Jones, D. L., & Golyshin, P. N. (2010). Effect of the earthworms *Lumbricus terrestris* and *Aporrectodea caliginosa* on bacterial diversity in soil. *Microbial Ecology*, 59, 574-587.
- Neuhauser, E. F., & Callahan, C. A. (1990). Growth and reproduction of the earthworm *Eisenia fetida* exposed to sublethal concentrations of organic chemicals. *Soil Biology and Biochemistry*, 22(2), 175-179.
- Nunes, M. E. T., Daam, M. A., & Espindola, E. L. G. (2016). Survival, morphology and reproduction of *Eisenia andrei* (Annelida, Oligochaeta) as affected by Vertimec® 18 EC (ai abamectin) in tests performed under tropical conditions. *Applied Soil Ecology*, 100, 18-26.
- Oluah, M. S., Obiezue, R. N. N., Ochulor, A. J., & Onuoha, E. (2010). Toxicity and histopathological effect of atrazine (herbicide) on the earthworm *Nsukkadrius mbae* under laboratory conditions. *Animal Research International*, 7(3), 1287-1293.
- Owagboriaye, F., Mesnage, R., Dedede, G., Adegboyega, T., Aladesida, A., Adeleke, M., Owa, S., & Antoniou, M. N. (2021). Impacts of a glyphosate-based herbicide on the gut microbiome of three earthworm species (*Alma millsoni*, *Eudrilus eugeniae* and *Libyodrilus violaceus*): a pilot study. *Toxicology Reports*, 8, 753-758.
- Panda, S., & Sahu, S. K. (2004). Recovery of acetylcholine esterase activity of *Drawida willsi* (Oligochaeta) following application of three pesticides to soil. *Chemosphere*, 55(2), 283-290.
- Pawar, S. S., & Ahmad, S. (2014). Filter paper contact test method for estimation of toxic effect of chlorpyrifos on earthworm, *Eisenia fetida*. *International Research Journal of Science Engineering*, 2(1), 23-25.
- Qiao, Z., Li, P., Tan, J., Peng, C., Zhang, F., Zhang, W., & Jiang, X. (2022). Oxidative stress and detoxification mechanisms of earthworms (*Eisenia fetida*) after exposure to flupyradifurone in a soil-earthworm system. *Journal of Environmental Management*, 322, 115989.
- Rani, L., Thapa, K., Kanojia, N., Sharma, N., Singh, S., Grewal, A. S., Srivastav, A. L., & Kaushal, J. (2021). An extensive review on the consequences of chemical pesticides on human health and environment. *Journal of Cleaner Production*, 283, 124657.
- Rao, J. V., & Kavitha, P. (2004). Toxicity of azodrin on the morphology and acetylcholinesterase activity of the earthworm *Eisenia foetida*. *Environmental Research*, 96(3), 323-327.
- Reddy, A. A., Reddy, M., & Mathur, V. (2024). Pesticide Use, Regulation, and Policies in Indian Agriculture. *Sustainability*, 16(17), 7839.
- Reddy, N. C., & Rao, J. V. (2008). Biological response of earthworm, *Eisenia foetida* (Savigny) to an organophosphorous pesticide, profenofos. *Ecotoxicology and Environmental Safety*, 71(2), 574-582.
- Reinecke, S. A., & Reinecke, A. J. (2007). The impact of organophosphate pesticides in orchards on earthworms in the Western Cape, South Africa. *Ecotoxicology and Environmental Safety*, 66(2), 244-251.

- Ribera, D., Narbonne, J. F., Arnaud, C., & Saint-Denis, M. (2001). Biochemical responses of the earthworm *Eisenia fetida andrei* exposed to contaminated artificial soil, effects of carbaryl. *Soil Biology and Biochemistry*, 33(7-8), 1123-1130.
- Robidoux, P. Y., Svendsen, C., Caumartin, J., Hawari, J., Ampleman, G., Thiboutot, S., Weeks, J. M., & Sunahara, G. I. (2000). Chronic toxicity of energetic compounds in soil determined using the earthworm (*Eisenia andrei*) reproduction test. *Environmental Toxicology and Chemistry: An International Journal*, 19(7), 1764-1773.
- Rodriguez-Campos, J., Dendooven, L., Alvarez-Bernal, D., & Contreras-Ramos, S. M. (2014). Potential of earthworms to accelerate removal of organic contaminants from soil: a review. *Applied Soil Ecology*, 79, 10-25.
- Römbke, J., Garcia, M. V., & Scheffczyk, A. (2007). Effects of the fungicide benomyl on earthworms in laboratory tests under tropical and temperate conditions. *Archives of Environmental Contamination and Toxicology*, 53, 590-598.
- Shi, Y., Shi, Y., Wang, X., Lu, Y., & Yan, S. (2007). Comparative effects of lindane and deltamethrin on mortality, growth, and cellulase activity in earthworms (*Eisenia fetida*). *Pesticide Biochemistry and Physiology*, 89(1), 31-38.
- Singh, S., Singh, J., & Vig, A. P. (2016). Earthworm as ecological engineers to change the physico-chemical properties of soil: soil vs vermicast. *Ecological Engineering*, 90, 1-5.
- Singh, S., Tiwari, R. K., & Pandey, R. S. (2020). An insight into the impact of triazophos and deltamethrin pesticides as individual and in combination on oxidative stress and histopathological alterations in *Eudrilus eugeniae*. *Chemistry and Ecology*, 36(2), 155-173.
- Sinha, M. P., Srivastava, R., & Gupta, D. K. (2013). Earthworm biodiversity of Jharkhand: Taxonomic description. *The Bioscan*, 8(Supplement 1), 293-310.
- Sorour, J., & Larink, O. (2001). Toxic effects of benomyl on the ultrastructure during spermatogenesis of the earthworm *Eisenia fetida*. *Ecotoxicology and Environmental Safety*, 50(3), 180-188.
- Sujeeth, N. K., Aravindh, R., Thandeewaran, M., Angayarkanni, J., Rajasekar, A., Mythili, R., & Gnanadesigan, M. (2023). Toxicity analysis and biomarker response of Quinalphos Organophosphate Insecticide (QOI) on eco-friendly exotic *Eudrilus eugeniae* earthworm. *Environmental Monitoring and Assessment*, 195(2), 274.
- Taillebois, E., Cartereau, A., Jones, A. K., & Thany, S. H. (2018). Neonicotinoid insecticides mode of action on insect nicotinic acetylcholine receptors using binding studies. *Pesticide Biochemistry and Physiology*, 151, 59-66.
- Tao, J., Rong, W., Diao, X., & Zhou, H. (2018). Toxic responses of Sox2 gene in the regeneration of the earthworm *Eisenia foetida* exposed to Retnoic acid. *Comparative Biochemistry and Physiology Part C: Toxicology & Pharmacology*, 204, 106-112.
- Thompson, C. M., & Richardson, R. J. (2004). Anticholinesterase insecticides. In: Marrs, T. C., & Ballantyne, B. (Eds) *Pesticide toxicology and international regulation*. West Sussex, England: Wiley. p 89-127
- Tiwari, R. K., Singh, S., & Pandey, R. S. (2019). Assessment of acute toxicity and biochemical responses to chlorpyrifos, cypermethrin and their combination exposed earthworm, *Eudrilus eugeniae*. *Toxicology Reports*, 6, 288-297.
- Van Gestel, C. A. M. (1992). Validation of earthworm toxicity tests by comparison with field studies: a review of benomyl, carbendazim, carbofuran, and carbaryl. *Ecotoxicology and Environmental Safety*, 23(2), 221-236.
- Velki, M., & Ežimović, S. (2015). Changes in exposure temperature lead to changes in pesticide toxicity to earthworms: a preliminary study. *Environmental Toxicology and Pharmacology*, 40(3), 774-784.
- Velki, M., Hackenberger, B. K., Lončarić, Ž., & Hackenberger, D. K. (2014). Application of microcosmic system for assessment of insecticide effects on biomarker responses in ecologically different earthworm species. *Ecotoxicology and Environmental Safety*, 104, 110-119.
- Wang, J. H., Zhu, L. S., Liu, W., Wang, J., & Xie, H. (2012). Biochemical responses of earthworm (*Eisenia foetida*) to the pesticides chlorpyrifos and fenvalerate. *Toxicology Mechanisms and Methods*, 22(3), 236-241.
- Wang, J., Wang, J., Wang, G., Zhu, L., & Wang, J. (2016). DNA damage and oxidative stress induced by imidacloprid exposure in the earthworm *Eisenia fetida*. *Chemosphere*, 144, 510-517.
- Wang, K., Pang, S., Mu, X., Qi, S., Li, D., Cui, F., & Wang, C. (2015). Biological response of earthworm, *Eisenia fetida*, to five neonicotinoid insecticides. *Chemosphere*, 132, 120-126.
- Wang, K., Qi, S., Mu, X., Chai, T., Yang, Y., Wang, D., Li, D., Che, W., & Wang, C. (2015). Evaluation of the toxicity, AChE activity and DNA damage caused by imidacloprid on earthworms, *Eisenia fetida*. *Bulletin of Environmental Contamination and Toxicology*, 95, 475-480.
- Wang, Y., Wu, S., Chen, L., Wu, C., Yu, R., Wang, Q., & Zhao, X. (2012). Toxicity assessment of 45 pesticides to the epigeic earthworm *Eisenia fetida*. *Chemosphere*, 88(4), 484-491.
- Wu, R., Zhou, T., Wang, J., Wang, J., Du, Z., Li, B., Juhasz, A., & Zhu, L. (2021). Oxidative stress and DNA damage induced by trifloxystrobin on earthworms (*Eisenia fetida*) in two soils. *Science of The Total Environment*, 797, 149004.
- Xiao, N., Jing, B., Ge, F., & Liu, X. (2006). The fate of herbicide acetochlor and its toxicity to *Eisenia fetida* under laboratory conditions. *Chemosphere*, 62(8), 1366-1373.
- Xu, P., Diao, J., Liu, D., & Zhou, Z. (2011). Enantioselective bioaccumulation and toxic effects of metalaxyl in earthworm *Eisenia foetida*. *Chemosphere*, 83(8), 1074-1079.
- Yao, X., Zhang, F., Qiao, Z., Yu, H., Sun, S., Li, X., Zhang, J., & Jiang, X. (2020). Toxicity of thifluzamide in earthworm (*Eisenia fetida*). *Ecotoxicology and Environmental Safety*, 188, 109880.
- Yasmin, S., & D'Souza, D. (2007). Effect of pesticides on the reproductive output of *Eisenia fetida*. *Bulletin of Environmental Contamination and Toxicology*, 79, 529-532.
- Yasmin, S., & D'Souza, D. (2010). Effects of pesticides on the growth and reproduction of earthworm: a review. *Applied and Environmental soil science*, 2010(1), 678360.
- Yuan, Y., Teng, H., Zhang, T., Wang, D., Gu, H., & Lv, W. (2024). Toxicological effects induced by two carbamates on earthworms (*Eisenia fetida*): Acute toxicity, arrested regeneration and underlying mechanisms. *Ecotoxicology and Environmental Safety*, 269, 115824.
- Zhang, C., Zhu, L., Wang, J., Wang, J., Du, Z., Li, B., Zhou, T., Cheng, C., & Wang, Z. (2018). Evaluating subchronic toxicity of fluoxastrobin using earthworms (*Eisenia fetida*). *Science of the total environment*, 642, 567-573.
- Zhang, Q., Zhang, B., & Wang, C. (2014). Ecotoxicological effects on the earthworm *Eisenia fetida* following exposure to soil contaminated with imidacloprid. *Environmental Science and Pollution Research*, 21, 12345-12353.
- Zhang, Q., Zhu, L., Wang, J., Xie, H., Wang, J., Han, Y., & Yang, J. (2013). Oxidative stress and lipid peroxidation in the earthworm *Eisenia fetida* induced by low doses of fomesafen. *Environmental Science and Pollution Research*, 20, 201-208.
- Zhang, R., Zhou, Z., & Zhu, W. (2020). Evaluating the effects of the tebuconazole on the earthworm, *Eisenia fetida* by H-1 NMR-Based untargeted metabolomics and mRNA assay. *Ecotoxicology and Environmental Safety*, 194, 110370.
- Zhang, X., Wang, X., Liu, Y., Fang, K., & Liu, T. (2020). The toxic effects of sulfoxaflor induced in earthworms (*Eisenia fetida*) under effective concentrations. *International Journal of Environmental Research and Public Health*, 17(5), 1740.
- Zhao, S., Dong, J., Jeong, H. J., Okumura, K., & Ueda, H. (2018). Rapid detection of the neonicotinoid insecticide imidacloprid using a quenchbody assay. *Analytical and bioanalytical chemistry*, 410, 4219-4226.
- Zhou, Q. X., Zhang, Q. R., & Liang, J. D. (2006). Toxic effects of acetochlor and methamidophos on earthworm *Eisenia fetida* in phaeozem, northeast China. *Journal of Environmental Sciences*, 18(4), 741-745.
- Zhou, S. P., Duan, C. Q., Hui, F. U., Chen, Y. H., Wang, X. H., & Yu, Z. F. (2007). Toxicity assessment for chlorpyrifos-contaminated soil with three different earthworm test methods. *Journal of Environmental Sciences*, 19(7), 854-858.
- Zhu, L., Li, B., Wu, R., Li, W., Wang, J., Wang, J., Du, Z., Juhasz, A., & Zhu, L. (2020). Acute toxicity, oxidative stress and DNA damage of chlorpyrifos to earthworms (*Eisenia fetida*): The difference between artificial and natural soils. *Chemosphere*, 255, 126982.
- Zoran, M. J., Heppner, T. J., & Drewes, C. D. (1986). Teratogenic effects of the fungicide benomyl on posterior segmental regeneration in the earthworm, *Eisenia fetida*. *Pesticide science*, 17(6), 641-652.